






Human Cathepsin D, CATH-D GENLISA™ ELISA

REF : KBH4668

Ver 2.1


RUO

Enzyme Immunoassay for the Quantitative Determination of Human Cathepsin D, CATH-D in serum, plasma and other biological samples.

| | | | |
|---|------------------------------|---|---------------------------------------|
| RUO | For Research Use Only | REF | Catalog Number |
|  | Store At | LOT | Batch Code |
|  | Manufactured By |  | Biological Risk |
|  | Expiry Date |  | Consult Operating Instructions |

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 96 tests

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Introduction:

The GENLISA™ ELISA kits are used for assessing the specific biomarker in samples analytes which may be serum, plasma and cell culture supernatant as validated with the kit. The kit employs a sandwich ELISA technique which leads to a higher specificity and increased sensitivity compared to conventional competitive ELISA kits which employ only one antibody. Double antibodies are used in this kit.

Intended Use:

The Human Cathepsin D, CATH-D GENLISA™ ELISA kit is used as an analytical tool for quantitative determination of Human Cathepsin D, CATH-D in serum, plasma and other biological samples.

Principle:

The method employs sandwich ELISA technique. Human Cathepsin D monoclonal antibodies are pre-coated onto microwells. Samples and standards are pipetted into microwells and Human Cathepsin D, CATH-D present in the sample are bound by the antibodies. Biotin labeled CATH-D antibody is added and followed by Streptavidin-HRP is pipetted and incubated to form a complex. After washing microwells in order to remove any non-specific binding, the substrate solution (TMB) is added to microwells and color develops proportionally to the amount of Human Cathepsin D, CATH-D in the sample. Color development is then stopped by addition of stop solution. Absorbance is measured at 450 nm.

Materials Provided:

1. Anti-Human CATH-D Antibody Coated Microtiter Plate (8 x 12 wells) – 1 no
2. Human CATH-D Standard (concentrated, 80 ng/ml) – 0.5 ml
3. Biotinylated CATH-D Antibody – 1 ml
4. Streptavidin:HRP Conjugate – 6 ml
5. Standard Diluent – 3 ml
6. (20X) Wash Buffer – 25 ml
7. TMB Substrate – 12 ml
8. Stop Solution – 12 ml
9. Instruction Manual

Materials to be provided by the End-User:

1. Microtiter Plate Reader able to measure absorbance at 450 nm.
2. Adjustable pipettes and multichannel pipettor to measure volumes ranging from 25 ul to 1000 ul
3. Deionized (DI) water
4. Wash bottle or automated microplate washer
5. Graph paper or software for data analysis
6. Timer
7. Absorbent Paper

Handling/Storage:

1. All reagents should be stored as indicated on the component label.
2. All the reagents and wash solutions should be used within 12 months from manufacturing date.
3. Before using, bring all components to room temperature (18-25°C). Upon assay completion ensure all components of the kit are returned to appropriate storage conditions.
4. The Substrate is light-sensitive and should be protected from direct sunlight or UV sources.

Health Hazard Warnings:

1. Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin.
2. For Research Use Only.



Sample Preparation and Storage:

Specimens should be clear and non-hemolyzed. Samples should be run at a number of dilutions to ensure accurate quantitation.

1. Extract as soon as possible after specimen collection as per relevant procedure. The samples should be tested as soon as possible after the extraction. Alternately the extracted samples can be kept in -20°C. Avoid repeated freeze-thaw cycles.
2. **Serum-** Coagulate at room temperature for 10-20 minutes; centrifuge for 20-min at 2000-3000 rpm. Remove the supernatant. If precipitation appears, recentrifuge.
3. **Plasma-** Use EDTA or citrate plasma as an anticoagulant, mix for 10-20 minutes; centrifuge for 15-min at 2000-3000 rpm. Remove the supernatant carefully. If precipitation appears, recentrifuge.
4. **Urine-** Collect urine in a sterile container, centrifuge for 20-min at 2000-3000 rpm. Remove the supernatant. If precipitation appears, recentrifuge.
5. **Cell Culture Supernatant-** Collect sample in a sterile container. Centrifuge for 20-mins at 2000-3000 rpm. Remove the supernatant carefully. When examining the components within the cell, dilute cell suspension with PBS (pH 7.2-7.4), if cell concentration is greater than 1 million/ml. Damage the cells by repeated freeze-thaw cycles to release intracellular components. Centrifuge for 20-min at 2000-3000 rpm. If precipitation appears, centrifuge again.
6. **Tissue Samples-** Rinse tissues in PBS (pH 7.4) to remove excess blood thoroughly and weigh before homogenization. Mince tissues and homogenize them in PBS (pH7.4) with a glass homogenizer on ice. Thaw at 2-8°C or freeze at -20°C. Centrifuge at 2000-3000 RPM for approximately 20 minutes and collect the supernatant carefully.

Note: Grossly hemolyzed samples are not suitable for use in this assay.

Reagent Preparation (all reagents should be diluted immediately prior to use):

1. Label any aliquots made with the kit Lot No and Expiration date and store it at appropriate conditions mentioned.
2. Bring all reagents to Room temperature before use.
3. To make **Wash Buffer (1X)**; dilute **25 ml of 20X Wash Buffer in 475 ml of DI water**.
4. **Standards Preparation:** Dilute 120 ul of original **Standard (80 ng/ml)** with 120 ul of standard diluent to generate a **40 ng/ml Standard stock solution**. Keep the standard for 15 mins with gentle agitation before making further dilutions. Prepare the **Standards** by serially diluting the standard stock solution as per the below table.

| Standard Concentration | Standard Vial | Dilution Particulars |
|------------------------|-------------------|---|
| 80 ng/ml | Original Standard | Original Standard provided in the Kit |
| 40 ng/ml | Standard No.5 | 120 ul Standard Provided (80 ng/ml) + 120 ul Standard Diluent |
| 20 ng/ml | Standard No.4 | 120 ul Standard No.5 + 120 ul Standard Diluent |
| 10 ng/ml | Standard No.3 | 120 ul Standard No.4 + 120 ul Standard Diluent |
| 5 ng/ml | Standard No.2 | 120 ul Standard No.3 + 120 ul Standard Diluent |
| 2.5 ng/ml | Standard No.1 | 120 ul Standard No.2 + 120 ul Standard Diluent |

Procedural Notes:

1. In order to achieve good assay reproducibility and sensitivity, proper washing of the plates to remove excess un-reacted reagents is essential.
2. High Dose Hook Effect may be observed in samples with very high concentrations of Human Cathepsin D, CATH-D. High Dose Hook Effect is due to excess of antibody for very high concentrations of Human Cathepsin D, CATH-D present in the sample.
3. Human Cathepsin D, CATH-D concentration of the undiluted sample is less than the diluted sample, this may be indicative of the Hook Effect.
4. Avoid assay of Samples containing sodium azide (NaN₃), as it could destroy the HRP activity resulting in under-estimation of the amount of Human Cathepsin D, CATH-D.
5. It is recommended that all Standards and Samples be assayed in duplicates or triplicates.
6. Maintain a repetitive timing sequence from well to well for all the steps to ensure that the incubation timings are same for each well.

7. If the Substrate has a distinct blue color prior to use it may have been contaminated and use of such substrate can lead to compromise of the sensitivity of the assay.
8. The plates should be read within 30 minutes after adding the Stop Solution.
9. Make a work list in order to identify the location of Standards and Samples.

Assay Procedure:

1. It is strongly recommended that all Standards and Samples be run in duplicates or triplicates. A standard curve is required for each assay.
2. Add **50 ul prepared Standards** to respective standard wells.
3. Add **40 ul Samples** to respective sample wells.
4. Pipette **10 ul Biotinylated CATH-D Antibody** to respective sample wells.
Note: Do not add **Biotinylated CATH-D Antibody** to standard wells. The standards provided in the kit are pre-offered as a complex of the standard and the biotin antibody for ease-of-use.
5. Pipette **50 ul Streptavidin:HRP Conjugate** to all wells. Mix well.
6. Cover the plate with a sealer and incubate for **60 minutes at 37°C**.
7. Aspirate and wash plate 4 times with diluted **Wash Buffer (1X)** and blot residual buffer by firmly tapping plate upside down on absorbent paper. Wipe of any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
8. Pipette **100 ul TMB Substrate** to all wells.
9. Incubate the plate at **37°C for 10 minutes**. DO NOT SHAKE or else it may result in higher backgrounds and worse precision. Positive wells should turn bluish in color.
10. Pipette **100 ul of Stop Solution** to all wells. The wells should turn from blue to yellow in color.
11. Read the absorbance at 450 nm with a microplate within 10-15 minutes after addition of Stop solution.

Calculation of Results:

Determine the Mean Absorbance for each set of duplicate or triplicate Standards and Samples. Using Graph Paper, plot the average value (absorbance 450nm) of each standard on the Y-axis versus the corresponding concentration of the standards on the X-axis. Draw the best fit curve through the standard points. To determine the unknown Human Cathepsin D, CATH-D concentrations, find the unknown's Mean Absorbance value on the Y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the X-axis and read the Human Cathepsin D, CATH-D Concentration.

If samples were diluted, multiply by the appropriate dilution factor. Software which is able to generate a cubic spline curve-fit is best recommended for automated results.

Note:

It is recommended to repeat the assay at a different dilution factor in the following cases:
- If the sample absorbance value is below the first standard.

Quality Control:

It is recommended that for each laboratory assay appropriate quality control samples in each run to be used to ensure that all reagents and procedures are correct.

Performance Characteristics of the Kit:

This kit has been validated. Please view the details herein below.

Standard Calibration Range:

2.5 ng/ml – 40 ng/ml

Sensitivity:**Limit Of Quantification:**

It is defined as the lowest detectable concentration that can be determined with an acceptable repeatability and the LOQ was found to be 2.1 ng/ml.

Specificity:

The antibodies used in this kit are monoclonal antibodies specific for Human Cathepsin D.

Precision:

Intra-Assay Precision: 3 samples (n=3) with low, middle and high concentration of Human Cathepsin D were tested in triplicate respectively. The Intra-Assay was found to be <15%

Inter-Assay Precision: 3 samples (n=3) with low, middle and high concentration of Human Cathepsin D were tested in triplicate on two plates respectively on two consecutive days. The Inter-Assay was found to be <18%.

The Cumulative Variance % was calculated as $CV (\%) = SD/mean \times 100$ [SD=standard deviation]

Safety Precautions:

- **This kit is For Research Use only.** Follow the working instructions carefully.
- The expiration dates stated on the kit are to be observed. The same relates to the stability stated for reagents
- Do not use or mix reagents from different lots.
- Do not use reagents from other manufacturers.
- Avoid time shift during pipetting of reagents.
- All reagents should be kept in the original shipping container.
- Some of the reagents contain small amount of sodium azide (< 0.1 % w/w) as preservative. They must not be swallowed or allowed to come into contact with skin or mucosa.
- Source materials maybe derived from Human body fluids or organs used in the preparation of this kit were tested and found negative for HBsAg and HIV as well as for HCV antibodies. However, no known test guarantees the absence of such viral agents. Therefore, handle all components and all patient samples as if potentially hazardous.
- Since the kit contains potentially hazardous materials, the following precautions should be observed
 - Do not smoke, eat or drink while handling kit material
 - Always use protective gloves
 - Never pipette material by mouth
 - Wipe up spills promptly, washing the affected surface thoroughly with a decontaminant.
- In any case GLP should be applied with all general and individual regulations to the use of this kit.

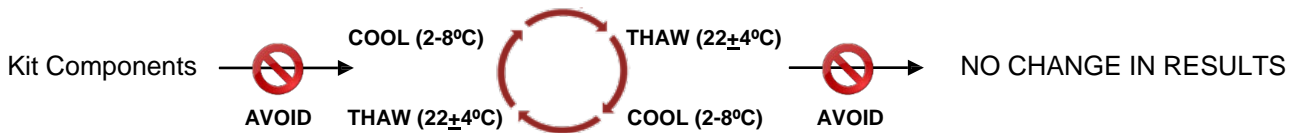


SCHEMATIC ASSAY PROCEDURE

1. Remove all components, 30 minutes before adding into the assay plate.



2. Avoid repeated cool-thaw of the components as there will be a loss of activity and this can affect the results.



3. Pipette **50 ul prepared Standards** into respective Standard wells.

4. Pipette **40 ul Samples** into the respective wells.

5. Pipette **10 ul Biotinylated Antibody** to the sample wells only. Note: Do not add Biotinylated Antibody to standard well because the Standard Solution contains the biotinylated antibody.

6. Pipette **50 ul Streptavidin:HRP Conjugate** to all wells.

7. Cover plate and incubate for at 37°C.

8. Aspirate and wash wells 4 times with **Wash Buffer (1X)**.

9. Pipette **100 ul TMB Substrate** to all wells.

10. Cover plate and incubate for at 37°C.

11. Pipette **100 ul Stop Solution** in all wells.

12. Read absorbance at 450nm with a microplate reader within of stopping reaction.

Typical Example of a Work List

| Well # | Contents | Absorbance at 450nm | Mean Absorbance | Interpolated Concentration |
|----------|--------------------------------|---------------------|-----------------|----------------------------|
| 1A 2A | Zero Std Zero Std | | | |
| 1B 2B | Standard No.1 Standard No.1 | | | |
| 1C 2C | Standard No.2 Standard No.2 | | | |
| 1D 2D | Standard No.3 Standard No.3 | | | |
| 1E 2E | Standard No.4 Standard No.4 | | | |
| 1F 2F | Standard No.5 Standard No.5 | | | |
| 1G 2G | Sample | | | |
| 1H 2H | Sample | | | |
| 3A 4A | Sample | | | |
| 3B 4B | Sample | | | |

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SYMBOLS KEY

| | |
|---|--------------------------------------|
|  | Coated Microtiter Plate (8x12 wells) |
|  | Standard |
|  | Biotinylated Antibody |
|  | Conjugate Horseradish Peroxidase |
|  | Standard Diluent |
|  | (20X) Wash Buffer |
|  | TMB Substrate |
|  | Stop Solution |
|  | Consult Instructions for Use |
|  | Catalog Number |
|  | Expiration Date |
|  | Storage Temperature |